### **STANDARD Q**

# COVID-19 Ag

STANDARD™ Q COVID-19 Ag Test

PLEASE READ CAREFULLY BEFORE YOU PERFORM THE TEST

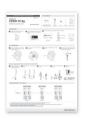
**SD BIOSENSOR** 

The device is intended for use in patients within 7 days of symptom onset.

# Test device (individually in a foil pouch with desiccant) Extraction buffer tube Nozzle cap Sterile swab Instructions for use

### **PREPARATION**

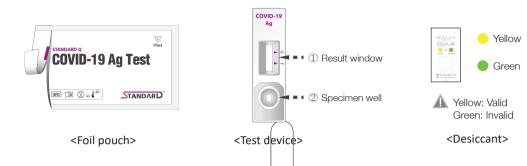
Carefully read instructions for using the STANDARD Q COVID-19 Ag Test.



2 Check the expiry date at the back of the foil pouch. Do not use the kit, if expiry date has passed.



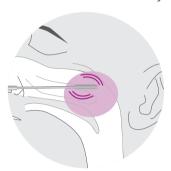
3 Check the test device and the desiccant pack in the foil pouch.



### **COLLECTION OF SPECIMEN**

### [Nasopharyngeal swab]

Insert a sterile swab into the nostril of the patient, swab over the surface of the posterior nasopharynx. Withdraw the sterile swab from the nasal cavity.



2 Insert the swab into an extraction buffer tube. While squeezing the buffer tube, stir the swab more than 5 times.



Remove the swab while squeezing the sides of the tube to extract the liquid from the swab.

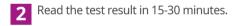


4 Press the nozzle cap tightly onto the tube.



### **ANALYSIS OF SPECIMEN**

1 Apply 3 drops of extracted specimen to the specimen well of the test device.



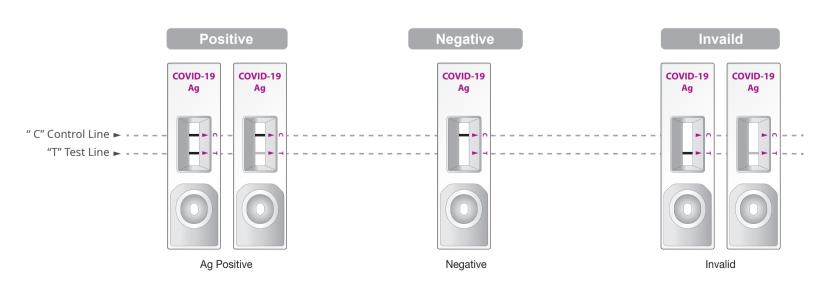






Do not read test results after 30 minutes.
 It may give false results.

### **INTERPRETATION OF TEST RESULT**



- 1. A colored band will appear in the top section of the result window to show that the test is working properly. This band is control line (C).
- 2. A colored band will appear in the lower section of the result window. This band is test line of SARS-CoV-2 antigen (T).
- 3. Even if the control line is faint, or the test line isn't uniform, the test should be considered to be performed properly and the test result should be interpreted as a positive result.
- \* The presence of any line no matter how faint the result is considered positive.
- \* Positive results should be considered in conjunction with the clinical history and other data available.

## **SD BIOSENSOR**

### **EXPLANATION AND SUMMARY**

### [Introduction]

. Coronavirus is a single-stranded positive-sense RNA virus with an envelope of about 80 to 120 nm in diameter. Its genetic material is the largest of all RNA viruses and is an important pathogen of many domestic animals, pets, and human diseases. It can cause a variety of acute and chronic diseases. Common signs of a person infected with a coronavirus include respiratory symptoms, fever, cough, shortness of breath, and dyspnea. In more severe cases, infection can cause pneumonia, severe acute respiratory syndrome, kidney failure, and even death. The 2019 new coronavirus, or "SARS-CoV-2 (COVID-19)", was discovered because of Wuhan Viral Pneumonia cases in 2019, and was named by the World Health Organization on January 12, 2020, confirming that it can cause colds and the Middle East Respiratory Syndrome (MERS) and more serious diseases such as acute respiratory syndrome (SARS). This kit is helpful for the auxiliary diagnosis of coronavirus infection. The test results are for clinical reference only and cannot be used as a basis for confirming or excluding cases alone.

### [Intended use]

STANDARD Q COVID-19 Ag Test is a rapid chromatographic immunoassay for the qualitative detection of specific antigens to SARS-CoV-2 present in human nasopharynx. This test is for administration by healthcare workers and labs only, as an aid to early diagnosis of SARS-CoV-2 infection in patient with clinical symptoms with SARS-CoV-2 infection. It provides only an initial screening test result. This product is strictly for medical professional use only and not intended for personal use. The administration of the test and the interpretation of the results should be done by a trained health professional. The result of this test should not be the sole basis for the diagnosis; confirmatory testing is required.

**[Test principle]**STANDARD Q COVID-19 Ag Test has two pre-coated lines, "C" Control line, "T" Test line on the surface of the nitrocellulose membrane. Both the control line and test line in the result window are not visible before applying any specimens. Mouse monoclonal anti-SARS-CoV-2 antibody is coated on the test line region and mouse monoclonal anti-Chicken IgY antibody is coated on the control line region. Mouse monoclonal anti-SARS-CoV-2 antibody conjugated with color particles are used as detectors for SARS-CoV-2 antigen device. During the test, SARS-CoV-2 antigen in the specimen interact with monoclonal anti-SARS-CoV-2 antibody conjugated with color particles making antigen-antibody color particle complex. This complex migrates on the membrane via capillary particles making antigen-antibody color particle compiex. Inis compiex migrates on the membrane via capillary action until the test line, where it will be captured by the mouse monoclonal anti-ARS-CoV-2 antibody. A colored test line would be visible in the result window if SARS-CoV-2 antigens are present in the specimen. The intensity of colored test line will vary depending upon the amount of SARS-CoV-2 antigen present in the specimen. If SARS-CoV-2 antigens are not present in the specimen, then no color appears in the test line. The control line is used for procedural control, and should always appear if the test procedure is performed properly and the test reagents

### [Kit contents]

① Test device (individually in a foil pouch with desiccant) ② Extraction buffer tube ③ Nozzle cap ④ Sterile swab ⑤ Instructions for use

### KIT STORAGE AND STABILITY

Store the kit at  $2-30^{\circ}$ C /  $36-86^{\circ}$ F out of direct sunlight. Kit materials are stable until the expiration date printed on the outer box. Do not freeze the kit.

### WARNINGS AND PRECAUTIONS

- Do not re-use the test kit. Do not use the test kit if the pouch is damaged or the seal is broken.
- Do not use the extraction buffer tube of another lot. Do not smoke, drink or eat while handling specimen.
- Wear personal protective equipment, such as gloves and lab coats when handling kit reagents. Wash hands thoroughly after the tests are done.
- Clean up spills thoroughly using an appropriate disinfectant.
- Handle all specimens as if they contain infectious agents.

  Observe established precautions against microbiological hazards throughout testing procedures.
- Dispose of all specimens and materials used to perform the test as bio-hazard waste. Laboratory chemical and biohazard wastes must be handled and discarded in accordance with all local, state, and national regulations.
   Desiccant in foil pouch is to absorb moisture and keep humidity from affecting products. If the moisture
- indicating desiccant beads change from yellow to green, the test device in the pouch should be discarded.

### SPECIMEN COLLECTION AND PREPARATION

- To collect a nasopharyngeal swab specimen, insert a sterile swab into the nostril of the patient, reaching the surface of the posterior nasopharynx.
- Using gentle rotation, push the swab until resistance is met at the level of the turbinate
- Rotate the swab a few times against the nasopharyngeal wall. Remove the swab from the nostril carefully.

- Specimen should be tested as soon as possible after collection.

  Once the specimen has been placed in the extraction buffer, the test needs to be performed within 1 hour at 15°C to 25°C and 4 hour at 2°C to 8°C.
- ens may be stored at room temperature for up to 1 hours or at 2-8  $^{\circ}$ C/ 36-46  $^{\circ}$ F for up to 4 hours prior to testing.

### PERFORMANCE CHARACTERISTICS

### [Clinical evaluation]

The sensitivity of the STANDARD Q COVID-19 Ag Test for rapid detection of SARS-CoV-2 antigen was established in prospective, multi institute, randomized, single-blinded study conducted at a trial site in Brazil and India during the 2020 SARS-CoV-2 pandemic. A total of 115 positive specimens were tested using the STANDARD Q COVID-19 Ag Test. These specimens consisted of nasopharyngeal swabs extracted directly from symptomatic patients. The specificity of STANDARD Q COVID-19 Ag Test was tested using 311 negative samples. The sensitivity and specificity of the STANDARD Q COVID-19 Ag Test was compared to a commercialized molecular assay.

### [Test sensitivity and specificity]

ved 96.52% of sensitivity and 99.68% of specificity.

Summary of the sensitivity and specificity of the STANDARD Q COVID-19 Ag Test compared to PCR.

		rcn			
		Positive	Negative	Total	
	Positive	111	1	112	
STANDARD Q COVID-19 Ag Test	Negative	4	310	314	
	Total	115	311	426	
Sensitivity		96.52% (111/1	15, 95% CI 91.3	3% - 99.04%)	
Specificity		99.68% (310/311, 95% CI 98.22 - 99.99%)			

### **ANALYTICAL PERFORMANCE**

Limit of Detection (LoD): The study used "SARS-CoV-2 (2019-nCOV) NCCP 43326/2020 / Korea" strain. The titer of cultured virus was confirmed by PCR. The cell is inactivated and spiked into Nasopharyngeal swab specimen The LoD is 1.25 X 10<sup>32</sup> TCID<sub>so</sub>/ml.

2019-nCoV Strain Tested	NCCP 43326/2020 / Korea								
Stock 2019-nCoV Titer		1 X 10 <sup>6.2</sup> TCID <sub>50</sub> /ml							
Dilution	1/10	1/100	1/200	1/400	1/800	1/1600	1/3200	1/6400	1/12800
Concentration in Dilution tested (TCID50/ml)	1 X 10 <sup>5.2</sup>	1 X 10 <sup>4.2</sup>	5 X 10 <sup>3.2</sup>	2.5 X 10 <sup>3.2</sup>	1.25 X 10 <sup>3.2</sup>	6.12 X10 <sup>2.2</sup>	3.06 X 10 <sup>2.2</sup>	1.53 X 10 <sup>2.2</sup>	7.1 X 10 <sup>1.2</sup>
Call rate of 5 replicates	100% (5/5)	100% (5/5)	100% (5/5)	100% (5/5)	100% (5/5)	80% (4/5)	0% (0/5)	0% (0/5)	0% (0/5)
Call rates of 20 replicates near cut-off	NA	NA	NA	NA	100% (20/20)	56% (14/20)	0% (0/20)	NA	NA
Lowest Concentration with Uniform Positivity per Analyte	1.25 X 10 <sup>32</sup> TCID <sub>50</sub> /ml								
Limit of Detection (LoD) per Virus Strain	1.25 X 10 <sup>3.2</sup> TCID <sub>sg</sub> /ml								

irus/Bacteria/ Parasite	Strain	Source/Specimen type	Concentration	Results	
SARS- coronavirus	Urbani	BEI/inactivated virus	3.5 ug/ml	POS	
MERS- coronavirus	Jeddah_1_2013	Bionote/recombinant protein	10 ug/ml	NEG	
	Type 1	Korea Bank for Pathogenic Viruses / live	3 X 10 <sup>5</sup> TCID <sub>50</sub> /ml	NEG	
	Type 3	National Culture Collection for pathogens / live	1.5 X 10 <sup>6</sup> TCID <sub>so</sub> /ml	NEG	
	Type 5	Korea Bank for Pathogenic Viruses / live	4 X 10 <sup>5</sup> TCID <sub>50</sub> /ml	NEG	
	Type 7	Korea Bank for Pathogenic Viruses / live	1.5 X 10 <sup>6</sup> TCID <sub>50</sub> /ml	NEG	
Adenovirus	Type 8	Korea Bank for Pathogenic Viruses / live	4 X 10 <sup>5</sup> TCID <sub>50</sub> /ml	NEG	
	Type 11	Korea Bank for Pathogenic Viruses / live	4 X 10 <sup>5</sup> TCID <sub>50</sub> /ml 4 X 10 <sup>5</sup>	NEG	
	Type 18	Korea Bank for Pathogenic Viruses / live	TCID <sub>50</sub> /ml	NEG	
	Type 23	Korea Bank for Pathogenic Viruses / live	4 X 10 <sup>5</sup> TCID <sub>50</sub> /ml	NEG	
	Type 55	Korea Bank for Pathogenic Viruses / live	4 X 10 <sup>5</sup> TCID <sub>50</sub> /ml	NEG	
	H1N1 Denver	ATCC/live virus	3 X 10 <sup>5</sup> TCID <sub>50</sub> /ml	NEG	
	H1N1 WS/33	ATCC/live virus	3 X 10 <sup>5</sup> TCID <sub>50</sub> /ml	NEG	
Influenza A	H1N1 Pdm-09	ATCC/live virus	3 X 10 <sup>5</sup> TCID <sub>50</sub> /ml	NEG	
	H1N1 New Caledonia	ATCC/live virus	3 X 10 <sup>5</sup> TCID <sub>50</sub> /ml	NEG	
	H1N1 New jersey	ATCC/live virus	3 X 10 <sup>5</sup> TCID <sub>50</sub> /ml	NEG	
	Nevada/03/2011	ATCC/live virus	3 X 10 <sup>5</sup> TCID <sub>50</sub> /ml	NEG	
Influenza B	B/Lee/40	ATCC/live virus	2.5 X 10 <sup>4</sup> TCID <sub>so</sub> /ml	NEG	
Dany!t-	B/Taiwan/2/62	ATCC/live virus	3 X 10 <sup>5</sup> TCID <sub>50</sub> /ml	NEG	
Respiratory syncytial virus Respiratory	Type A	ATCC/live virus	3 X 10 <sup>5</sup> TCID <sub>50</sub> /ml	NEG	
syncytial virus	Туре В	ATCC/live virus	3 X 10 <sup>5</sup> TCID <sub>50</sub> /ml	NEG	
Legionella	Bloomington-2	ATCC/live virus	5 X 10 <sup>4</sup> cells/ml	NEG	
pneumophila	Los Angeles-1	ATCC/live virus	5 X 10 <sup>4</sup> cells/ml	NEG	
	82A3105	ATCC/live virus	5 X 10 <sup>4</sup> cells/ml	NEG	
	K		5 X 10 <sup>4</sup> cells/ml	NEG	
Mycobacterium	Erdman	Vencei Univ. Cine stirete de la dele	5 X 10 <sup>4</sup> cells/ml	NEG	
tuberculosis	HN878 CDC1551	Yonsei Univ. / inactivated and filter	5 X 10 <sup>4</sup> cells/ml	NEG NEG	
	H37Rv		5 X 10° cells/ml	NEG	
	4752-98	ATCC/live	5 X 104 cells/ml	NEG	
Strantacaccii	[Maryland (D1)6B-17] 178 [Poland 23F-16]	ATCC/live	5 X 10 <sup>4</sup> cells/ml	NEG	
Streptococcus pneumonia	262 [CIP 104340]	ATCC/live	5 X 10 <sup>4</sup> cells/ml	NEG	
	Slovakia 14-10 [29055]	ATCC/live	5 X 10 <sup>4</sup> cells/ml	NEG	
Streptococcus pyrogens	Typing strain T1 [NCIB 11841, SF 130]	ATCC/live	5 X 10 <sup>4</sup> cells/ml	NEG	
17 -3	Mutant 22	ATCC/live	5 X 10 <sup>4</sup> cells/ml	NEG	
Mycoplasma pneumoniae	FH strain of Eaton Agent [NCTC 10119]	ATCC/live	5 X 10 <sup>4</sup> cells/ml	NEG	
	M129-B7	ATCC/live	5 X 10 <sup>4</sup> cells/ml	NEG	
Pooled human nasal wash - to represent diverse microbial flora in the human respiratory tract	NA	Bionote / Normal pooled human nasal wash from healthy employees SD biosensor / Normal pooled human nasal wash from healthy	NA	NEG	
	229E	employees Zeptomatrix/inactivaed	1 X 10 <sup>4.5</sup> TCID <sub>50</sub> /ml	NEG	
Coronavirus	OC43	Zeptomatrix/inactivaed	1 X 10 <sup>5</sup> TCID <sub>50</sub> /ml	NEG	
	NL63	Zeptomatrix/inactivaed	1 X 10 <sup>4</sup> TCID <sub>50</sub> /ml	NEG	
MERS-Coronavirus	Florida/USA-2_Saudi Arabia_2014	Zeptomatrix/inactivaed	4 X 10 <sup>4</sup> TCID <sub>50</sub> /ml	NEG	
Human etapneumovirus (hMPV) 3 Type B1	Peru2-2002	Zeptomatrix/inactivaed	1 X 10 <sup>5</sup> TCID <sub>50</sub> /ml	NEG	
Human Metapneumovirus (hMPV) 16 Type A1	IA10-2003	Zeptomatrix/inactivaed	1 X 10 <sup>5</sup> TCID <sub>50</sub> /ml	NEG	
	Type 1	Zeptomatrix/inactivaed	1 X 10 <sup>5</sup> TCID <sub>50</sub> /ml	NEG	
Parainfluenza	Type 2	Zeptomatrix/inactivaed	1 X 10 <sup>5</sup> TCID <sub>50</sub> /ml	NEG	
virus	Type 3	Zeptomatrix/inactivaed	1 X 10 <sup>5</sup> TCID <sub>50</sub> /ml	NEG	
	Type 4A	Zeptomatrix/inactivaed	1 X 105 TCID <sub>50</sub> /ml	NEG	

\*Human coronavirus HKU1 has not been tested. The % identity of the nucleocapsid protein sequence between HKU1 and SARS-CoV-2 is below 35%

Zeptomatrix/inactivaed

1 X 105 TCID../ml

Endogenous/Exogenous Interference Substances Studies: There was no interference for potential interfering substances listed below

Potential Interfering Substance	Concentration	Results
Respii	ratory Specimens	
Mucin: bovine submaxillary gland, type I-S	100 ug/ml	NEG
Blood (human), EDTA anticoagulated	5% (v/v)	NEG
Biotin	100 ug/ml	NEG
Nasa	l sprays or drops	
Neo-Synephrine (Phenylephrine)	10% (v/v)	NEG
Afrin Nasal Spray (Oxymetazoline)	10% (v/v)	NEG
Saline Nasal Spray	10% (v/v)	NEG
Homeopathi	c allergy relief medicine	
Homeopathic Zicam Allergy Relief Nasal Gel	5% (v/v)	NEG
Sodium Cromoglycate	20 mg/ml	NEG
Olopatadine Hydrochloride	10 mg/ml	NEG
An	nti-viral drugs	
Zanamivir (Influenza)	5 mg/ml	NEG
Oseltamivir (Influenza)	10 mg/ml	NEG
Artemether-lumefantrine (Malaria)	50 uM	NEG
Doxycycline hyclate (Malaria)	70 uM	NEG
Quinine (Malaria)	150 uM	NEG
Lamivudine (Retroviral medication)	1 mg/ml	NEG
Ribavirin (HCV)	1 mg/ml	NEG
Daclatasvir (HCV)	1 mg/ml	NEG

Anti-inflammatory medication				
Acetaminophen	199 uM	NEG		
Acetylsalicylic acid	3.62 mM	NEG		
Ibuprofen	2.425 mM	NEG		
	Antibiotic			
Mupirocin	10 mg/ml	NEG		
Tobramycin	5 ug/ml	NEG		
Erythromycin	81.6 uM	NEG		
Ciprofloxacin	30.2 uM	NEG		

Potential Interfering Substance	Concentration	Viral Strain Level (In multiples of LoD)	Results (Detected X/3)
Re	espiratory Specim	iens	
Mucin: bovine submaxillary gland, type I-S	100 ug/ml	SARS-CoV-2	POS
Blood (human), EDTA anticoagulated	5% (v/v)	cultured virus media 1/800 dilution	POS
Biotin	100 ug/ml	(1.25 X 10 <sup>3.2</sup> TCID <sub>50</sub> /ml)	POS
N	lasal sprays or dr	ops	'
Neo-Synephrine (Phenylephrine)	10% (v/v)	SARS-CoV-2	POS
Afrin Nasal Spray (Oxymetazoline)	10% (v/v)	cultured virus media	POS
Saline Nasal Spray	10% (v/v)	1/800 dilution (1.25 X 10 <sup>3.2</sup> TCID <sub>so</sub> /ml)	POS
Homeop	athic allergy relie	f medicine	
Homeopathic Zicam Allergy Relief Nasal Gel	5% (v/v)	SARS-CoV-2	POS
Sodium Cromoglycate	20 mg/ml	cultured virus media	POS
Olopatadine Hydrochloride	10 mg/ml	1/800 dilution (1.25 X 10 <sup>3.2</sup> TCID <sub>so</sub> /ml)	POS
	Anti-viral drugs	30	
Zanamivir (Influenza)	5 mg/ml		POS
Oseltamivir (Influenza)	10 mg/ml		POS
Artemether-lumefantrine (Malaria)	50 uM	SARS-CoV-2	POS
Doxycycline hyclate (Malaria)	70 uM	cultured virus media	POS
Quinine (Malaria)	150 uM	1/800 dilution	POS
Lamivudine (Retroviral medication)	1 mg/ml	(1.25 X 10 <sup>3.2</sup> TCID <sub>50</sub> /ml)	POS
Ribavirin (HCV)	1 mg/ml	1	POS
Daclatasvir (HCV)	1 mg/ml		POS
Anti-i	nflammatory med	dication	
Acetaminophen	199 uM	SARS-CoV-2	POS
Acetylsalicylic acid	3.62 mM	cultured virus media	POS
Ibuprofen	2.425 mM	1/800 dilution (1.25 X 10 <sup>3.2</sup> TCID <sub>50</sub> /ml)	POS
	Antibiotic	30	
Mupirocin	10 mg/ml	SARS-CoV-2	POS
Tobramycin	5 ug/ml	cultured virus media	POS
Erythromycin	81.6 uM	1/800 dilution	POS
Ciprofloxacin	30.2 uM	(1.25 X 10 <sup>3.2</sup> TCID <sub>50</sub> /ml)	POS

High-dose Hook Effect: SARS-CoV-2 cultured virus was spiked into specimen. SARS-CoV-2 cultured virus did

Specimen Type	Dilution	Concentration (TCID <sub>50</sub> /ml)	Result
	NEAT	1 X 10 <sup>6.2</sup>	POS
	1/10	1 X 10 <sup>5.2</sup>	POS
SARS-CoV-2 NCCP 43326/2020 / korea	1/100	1 X 10 <sup>4.2</sup>	POS
	1/200	5 X 10 <sup>3.2</sup>	POS
	1/400	2.5 X 10 <sup>3.2</sup>	POS
Inactivated virus cultured media	1/800	1.25 X 10 <sup>3.2</sup>	POS
	1/1600	6.12 X 10 <sup>2.2</sup>	POS
	1/3200	3.06 X 10 <sup>2.2</sup>	NEG
	1/6400	1.53 X 10 <sup>2.2</sup>	NEG
	1/12800	7.1 X 10 <sup>1.2</sup>	NEG

### LIMITATION OF TEST

- The test procedure, precautions and interpretation of results for this test must be followed strictly when testing. The test should be used for the detection of SARS-CoV-2 antigen in human nasopharyngeal swab specimens. Neither the quantitative value nor the rate of SARS-CoV-2 antigen concentration can be determined by this
- Failure to follow the test procedure and interpretation of test results may adversely affect test performance and/or produce invalid results.
- A negative test result may occur if the level of extracted antigen in a specimen is below the sensitivity of the test or if a poor quality specimen is obtained.

  For more accuracy of immune status, additional follow-up testing using other laboratory methods is recommended.
- The test result must always be evaluated with other data available to the physician.

  A negative result may occur if the concentration of antigen or antibody in a specimen is below the detection limit of the test or if the specimen was collected improperly, therefore a negative test result does not eliminate the possibility of SARS-CoV-2 infection, and should be confirmed by viral culture or an molecular assay or ELISA.
- Positive test results do not rule out co-infections with other pathogens. Negative test results are not intended to rule in other coronavirus infection except the SARS-CoV. Children tend to shed virus for longer periods of time than adults, which may result in differences in sensitivity
- between adults and children.

## **BIBLIOGRAPHY**

- Clinical management of severe acute respiratory infection when novel coronavirus(nCoV) infection is Diagnostic detection of Wuhan coronavirus 2019 by real-time RT-PCR.2020
- Diagnosis and treatment of pneumonia caused by new coronavirus (trial version 4) National Health

Information about sponsor

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